# BOSTON JNIVERSITY

# **Software for Differential Characterization of PTMs: Approaches in Data Acquisition and Processing** Jean L. Spencer, Vivek N. Bhatia, Amanuel Kehasse, Stephen A. Whelan, Christian F. Heckendorf,

## **Overview**

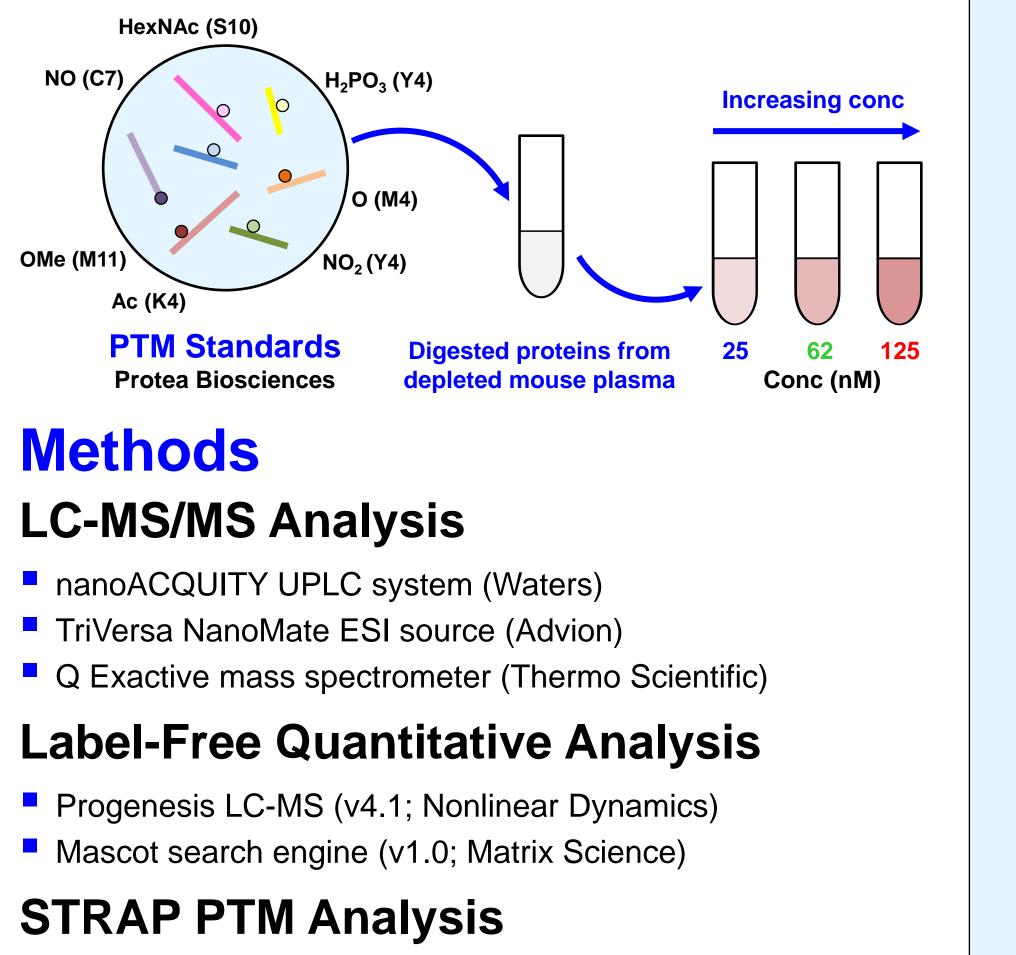
- Purpose: Investigate software that characterizes differential PTMs in complex data sets and determine the effects of data acquisition/processing parameters on its capability.
- Methods: In-house software (STRAP PTM) uses spectral counting and a novel scoring algorithm to collate and rank differential PTMs.
- Results: Trends in differential PTMs readily displayed and substantiated by quantitative analysis with results easily optimized by variation of acquisition/processing parameters.

### Introduction

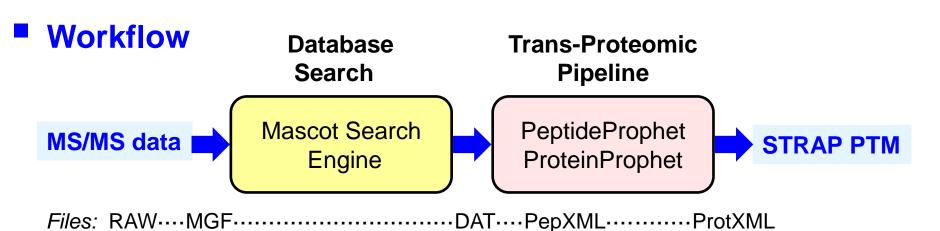
The identification of post-translational modifications (PTMs) across different states is critical for determining biomarkers and therapeutic targets in proteomics studies. Although software can identify peptides and by inference proteins, identification of PTMs is more challenging. Once PTMs are identified, few methods exist to analyze significant trends. We created **STRAP** PTM (Software Tool for Rapid Annotation of Proteins: Post-**Translational Modification edition)** to facilitate multi-sample comparison by collating and ranking PTMs. Here we explore the utility of STRAP PTM and the effects of data acquisition and processing parameters on its capability.

## **Materials**

### **PTM Peptide Standards in Plasma**



Software: STRAP PTM (v1.0 beta) freely available at http://www.bumc.bu.edu/cardiovascularproteomics/cpctools



### **Methods STRAP PTM Analysis** Setup Window - Enter protXML data files. - Select peptide probability cutoff. - Choose protein group overlap. - Download PTM database. Enter protein sequence database - Select PTM score factors. Execute analysis. PTM Scoring **PTM Score (S)**: Overall score for a specific PTM (m) on a specific site (i) of a specific protein (p) based on userselectable factors relevant to the system (max = 100) $= 100 \times Q_{mip} \times G_{mip} \times W_{mip} \times U_{mp}$ Occupancy Uniqueness Grouping **User-selectable factors** Quality (Q): Goodness of database search results for a specific PTM on a specific site of a specific protein (max = 1) $\langle \mathbf{P}_{mip} \rangle$ P = probability of modified peptides $\mathbf{Q}_{mip} = \frac{\langle \mathbf{P}^0 \rangle}{\langle \mathbf{P}^0 \rangle}$ $P^0$ = probability of unmodified peptides Probabilitv $\sim$ 0.2 - Q = $\frac{0.2/1}{1.0}$ = 0.2 $- Q = \frac{1.6/2}{1.0} = 0.8$ $\longrightarrow$ Better results Grouping (G): Variation of a specific PTM on a specific site of a specific protein across groups (max = 1) $\sigma_{mip}$ $\sigma$ = std dev of counts $G_{mip} = \frac{mip}{mip}$ max $\sigma$ = max $\sigma$ of all proteins $\max \sigma$ Group A Group B G = 2.1/2.1 = 1 G = 0/2.1 = 0 G = 0/2.1 = 0 G = 0/2.1 = 0**Occupancy (W):** Degree of modification of a specific site on a specific protein with a specific PTM (max = 1) $\mathbf{W}_{mip} = \frac{1}{\left(\sum_{m=1}^{M} \mathbf{N}_{mip}\right) + \left(\mathbf{N}^{0}_{ip}\right)}$ $(N^{0}_{in})$ N = modified peptide counts N<sup>0</sup> = unmodified peptide counts W = 1/4 = 0.25W = 2/4 = 0.50 Greater modification $\sim$ Uniqueness (U): Rarity of a specific PTM on a specific protein (max = 1) $\sum_{i=1}^{I} \mathbf{N}_{mip}$ I = total sites $\mathbf{U}_{mp} = \mathbf{1} - \frac{1}{\sum_{m=1}^{M} \sum_{i=1}^{I} \mathbf{N}_{mip}}$ M = total PTMs N = modified peptide counts U = 1 - 1/4 = 0.75 Less frequent U = 1 - 3/4 = 0.25

S <sub>mip</sub>
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# Results **STRAP PTM Results**

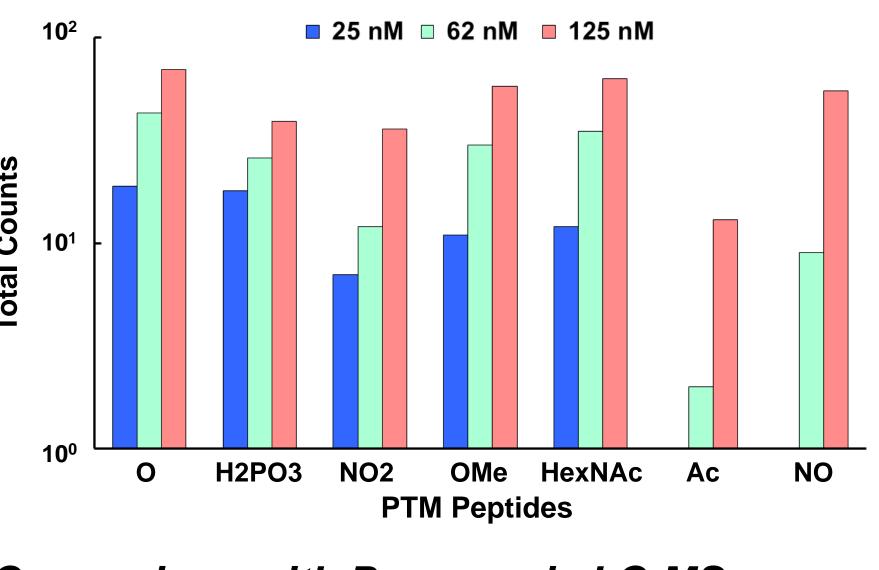
**PTM Scores** (top 7 protein

Peptide	РТМ	PTM Score QGW x 100	T 25 nM	otal Cou 62 nM	nts 125 nM	Modified Forms	Other Forms	Q	Scoring G	Factors W	U
PS-280	0	77.9	19	43	70	132	0	0.901	0.865	1.000	0
PS-500	HexNAc	73.0	12	35	63	110	0	0.844	0.866	1.000	0
PS-580	NO	71.1	0	9	55	64	21	0.944	1.000	0.753	0
PS-412	ОМе	67.6	11	30	58	99	0	0.844	0.801	1.000	0
PS-321	NO2	39.9	7	12	36	55	0	0.759	0.525	1.000	0
PS-301	H2PO3	35.5	18	26	39	83	1	1.000	0.359	0.988	0
PS-532	Ac	20.3	0	2	13	15	0	0.855	0.237	1.000	0

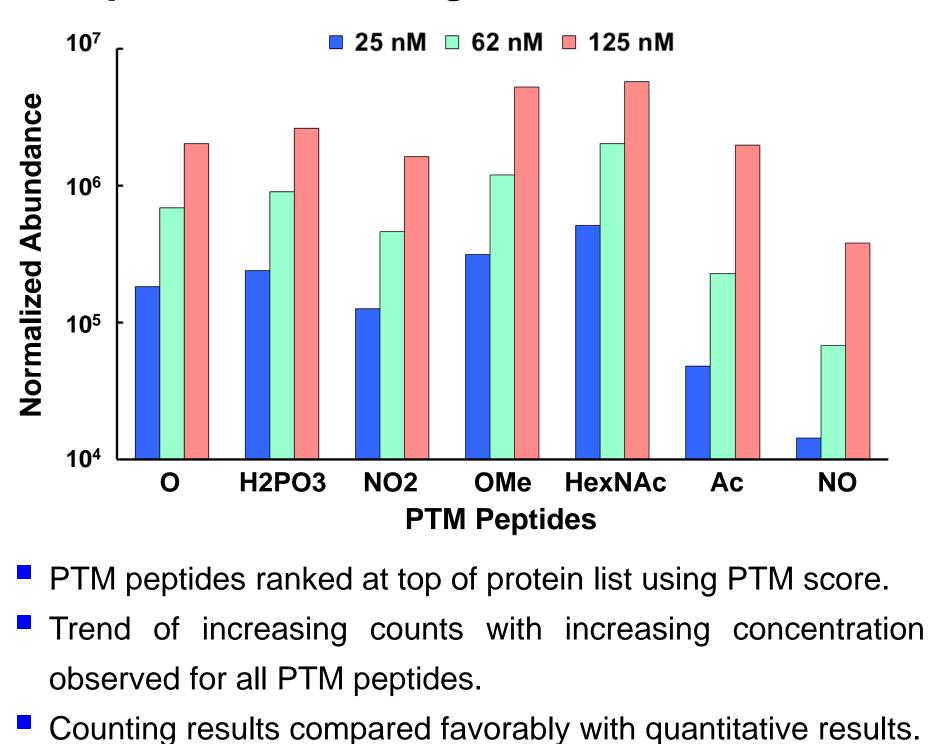
### PTM Map (location of counts at 25 nM, 62 nM, 125 nM)

Peptide	PTM (Site)
<b>PS-280</b>	O (M4)
<b>PS-301</b>	H2PO3 (Y4)
<b>PS-321</b>	NO2 (Y4)
<b>PS-412</b>	OMe (M11)
PS-500	HexNAc (S10)
<b>PS-532</b>	Ac (K4)
<b>PS-580</b>	NO (C7)

### PTM Counts



### Comparison with Progenesis LC-MS



# **Catherine E. Costello and Mark E. McComb**

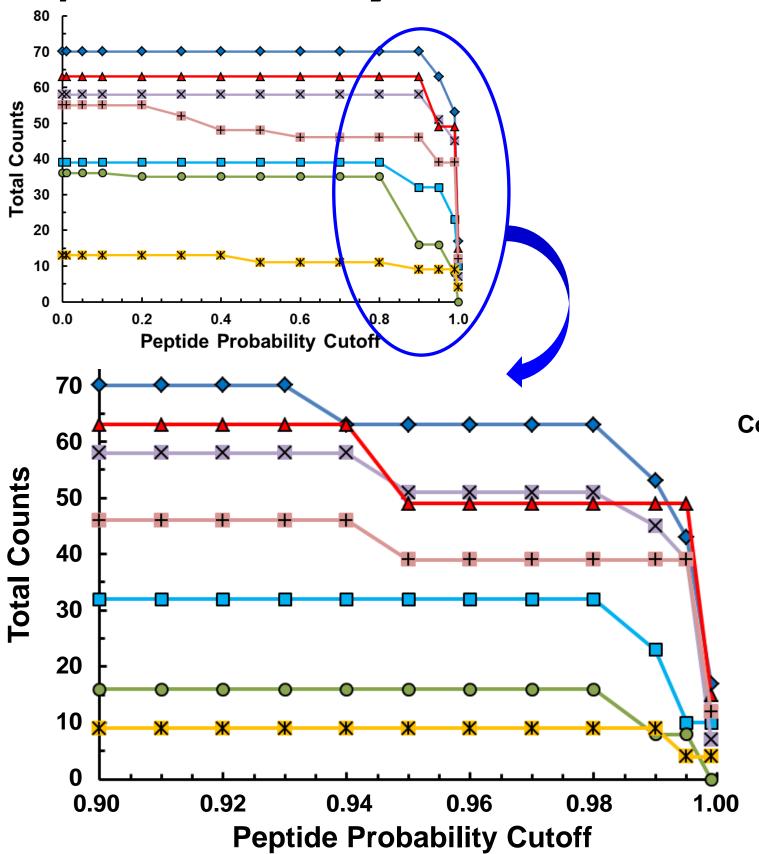
Cardiovascular Proteomics Center, Boston University School of Medicine, Boston, MA 02118

n hits)
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### Results

### Effect of STRAP PTM Settings Peptide Probability Cutoff



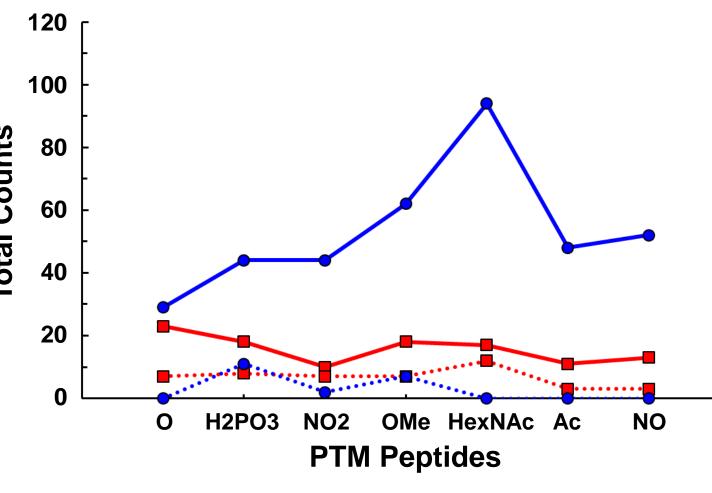
- Data quality improved by filtering out poorer search results with higher peptide probability cutoff.
- Better quality data balanced against falloff in counts (p > 0.99).

### Effect of Instrument Settings

Method	Mode	Description	DE (s)	IT (ms)	List	MSX	In
1	DD-MS2	Standard settings: DE on	4	60			8
2	DD-MS2	DE off		60			8
3	DD-MS2	List on; DE on	4	60	Yes		8
4	DD-MS2	List on; DE off		60	Yes		4
5	T-MS2	List on		60	Yes		
6	T-MS2	List on; IT short		20	Yes		
7	T-MS2	List on; IT short; multiplexing		20	Yes	7	

DD-MS2 = data-dependent MS/MS; T-MS2 = targeted MS/MS DE = dynamic exclusion; IT = injection time; List = inclusion list (PTM peptides) MSX = multiplexing; AGC = automatic gain control

### **DD-MS2 and Dynamic Exclusion**



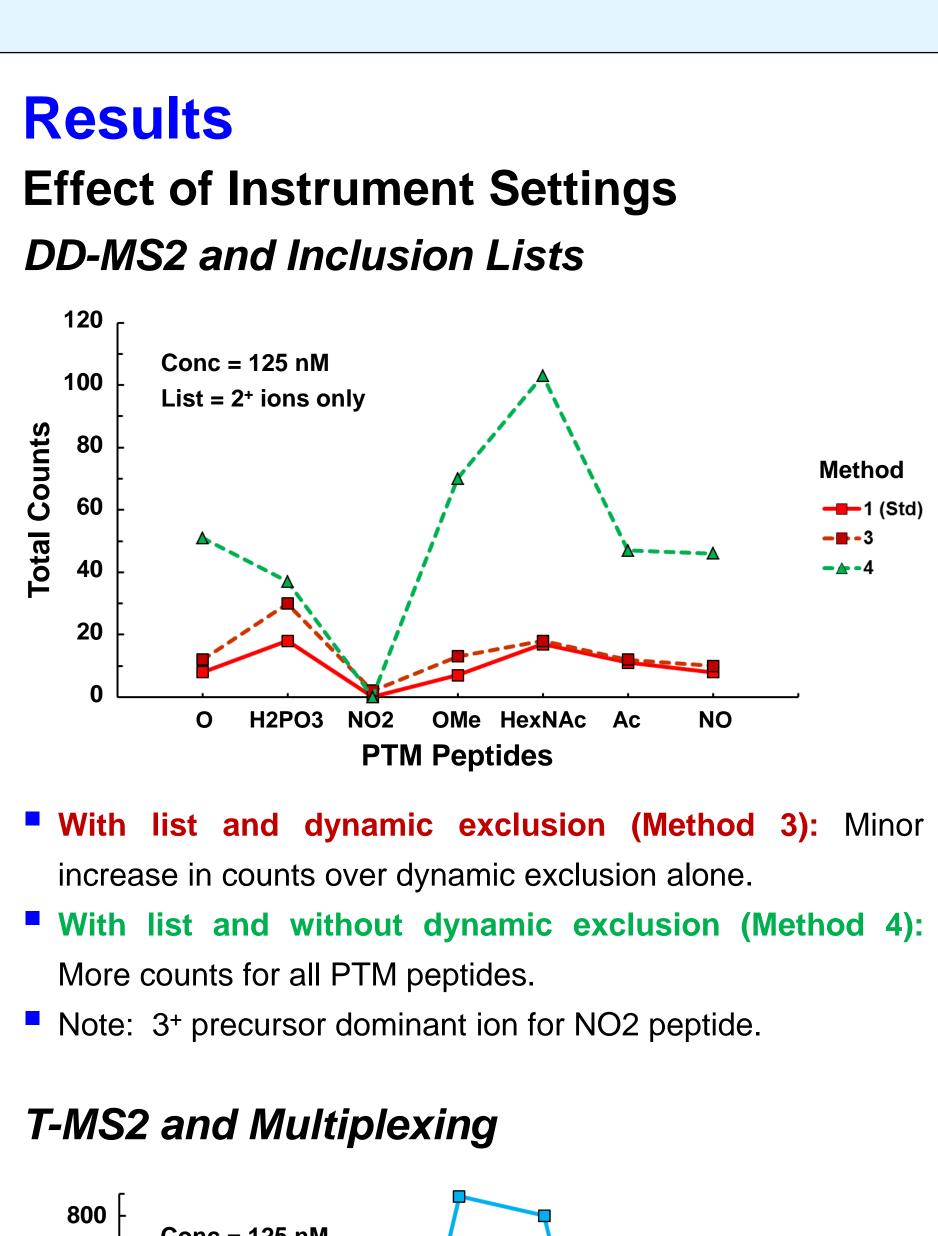
- With dynamic exclusion (Method 1): All PTM peptides detected at each concentration.
- Without dynamic exclusion (Method 2): More counts for all PTM peptides at highest concentration; 4 PTM peptides lost at lowest concentration (masked by higher peaks).

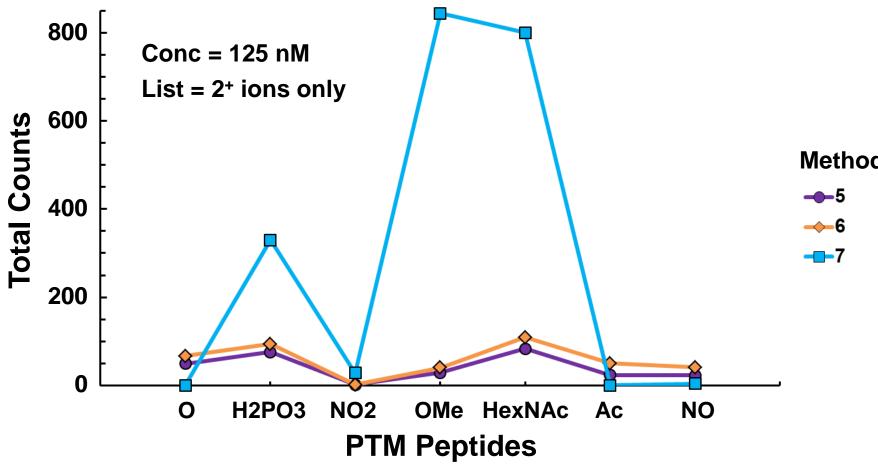
onc = 125 nM
<b>→</b> 0
- <b></b> H2PO3
NO2
–≍– OMe
HexNAc
<u>–*–</u> Ac
NO

itensity	AGC
<mark>.3 x 10<sup>4</sup></mark>	<mark>5.0 x 10<sup>5</sup></mark>
.3 x 10 <sup>4</sup>	5.0 x 10 <sup>5</sup>
.3 x 10 <sup>4</sup>	5.0 x 10 <sup>5</sup>
.2 x 10 <sup>5</sup>	5.0 x 10 <sup>5</sup>
	5.0 x 10 <sup>5</sup>
	3.0 x 10 <sup>6</sup>
	3.0 x 10 <sup>6</sup>

### Method —∎—1 (125 nM) ••**■**•1 (25 nM)

──2 (125 nM) •••••2 (25 nM)





- With shorter injection time (Method 6): More counts for all PTM peptides (higher duty cycle).
- With multiplexing and shorter injection time (Method 7): Inconsistent results from interference with other precursor ions.

# Conclusions

**STRAP PTM** is a powerful counting approach that:

- Collates and ranks differential PTMs in complex data sets.
- Generates results with trends substantiated by label-free, quantitative analysis.
- Allows removal of poorer quality data for improved analysis.
- Provides a rapid means to optimize instrument settings for best results.
- Represents a fast and easy tool for semi-quantitative results.

# Acknowledgments

- NIH-NHLBI contract HHSN268201000031C
- NIH grants P41 GM104603, R21 HL107993, S10 RR020946