

MicroRNA Expression Profiling Distinguish Normal From Neoplastic Urothelium

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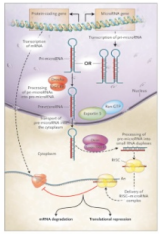
Introduction

The majority of studies, to date, on the genetic mutations that contribute to bladder cancer formation have focused on alterations to the genome and expression of protein coding genes.

Increasing evidence indicates, however, that dysregulation of microRNAs is associated with cancer and their expression profile can be correlated with disease pathogenesis and diagnosis.

The dysregulation of microRNAs has been observed in many cancers, including those in the breast, lung, colon, and pancreas.

In this report, we describe a procedure designed to identify the global expression pattern of microRNA expression in low and high grade bladder cancer. The goal is to define the microRNAs that can differentiate low and high grade bladder cancer from normal tissue for diagnostic and prognostic reasons.



Furthermore, recognition of microRNAs that are differentially expressed between the normal and tumor may help to identify those that are involved in bladder cancer and establish the basis to unravel their pathogenic role.

Patient Study

Tissue Specimens

Bladder tissue specimens were obtained from patients at Boston Medical. Institutional Review Board-approved informed consent for the collection of specimens was obtained from all patients. Seven of the tissue specimens were classified as high-grade, eight of the specimens were classified as low-grade, while ten of the specimens were normal urothelium. These fresh operative specimens were then put into five volumes of RNA later, set at room temperature for 1 hour, and stored at -80 C until RNA extraction.

cDNA Synthesis

Total RNA was prepared using the mir-VANA microRNA Isolation Kit (Ambion, Austin, TX, USA), and subsequently quantified using a spectrophotometer. Two micrograms of total RNA were reverse transcribed into first-strand cDNA using the QuantiMir Kit (System Biosciences, Mountain View, CA, USA), which tags and converts small RNAs into detectable cDNAs for qPCR.

Real-time quantitative PCR

Each qPCR reaction contained 5.0 ng of synthesized cDNA, 300 nM Universal reverse primer and 500nM microRNA-specific forward assay primer in a total reaction volume of 20ul. The sequence of the microRNA-specific forward primer was identical to the target microRNA being measured. The entire qPCR plate containing 96 assays (see microRNA assay panel) was subjected to standard real-time qPCR thermocycling and data collection. All reactions were performed in triplicate using an ABI 7900HT Sequence Detection System with PowerSYBR reagents (Applied Biosystems, Foster City, CA).

Method

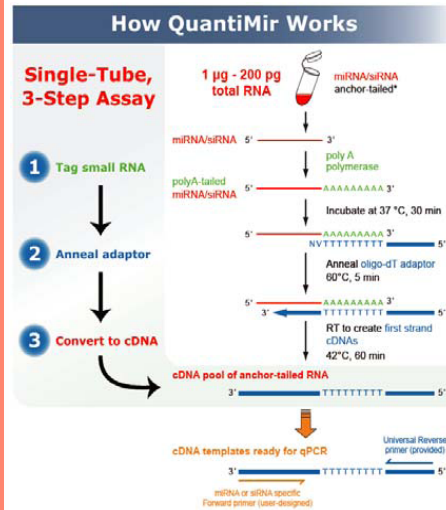


Plate Array Arrangement	
	1 2 3 4 5 6 7 8 9 10 11 12
A	miR.7 miR.92 miR.93 miR.9.1 miR.101.1 miR.103 miR.106.a miR.106.b miR.107 miR.106 miR.1.1
B	miR.122a miR.125a miR.125b miR.126 miR.128b miR.132 miR.133a miR.134 miR.135b miR.136 miR.137 miR.140 miR.142 miR.141 3p miR.143 miR.145 miR.146.a miR.149 miR.150 miR.151 miR.153 miR.154 miR.155 miR.15a
C	miR.15b miR.16 3p miR.17 3p miR.17a miR.181a miR.181b miR.181c miR.181d miR.183 miR.185 miR.186 miR.188 miR.189 miR.190 miR.191 miR.192 miR.194 miR.195 miR.196a miR.197 miR.198 3p miR.200 miR.201 miR.202 miR.203 miR.204 miR.205 miR.206 miR.21 miR.210
D	miR.214 miR.215 miR.217 miR.218 miR.219 miR.22 miR.488 miR.221 miR.222 miR.223 miR.224
E	miR.23a miR.24 miR.25 miR.26a miR.26b 27a+b miR.30c 29a+b+c 3p 5p miR.30a miR.30a miR.296 miR.296 miR.106

Statistical Method

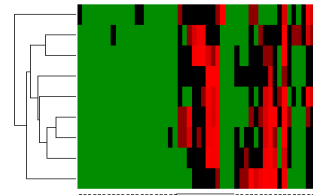
- Data transformed via DeltaCt: $2^{-(Ct_{target} - Ct_{control})}$
- Control miR: 106b

- Independent samples t-test run for HG miR vs. Normal miR
- Independent samples t-test run for LG miR vs. Normal miR
- Reject when $p < 0.05$ (i.e. these are miR with different means)
- Find non-overlapping miR from rejected lists, thus creating separate identifiers

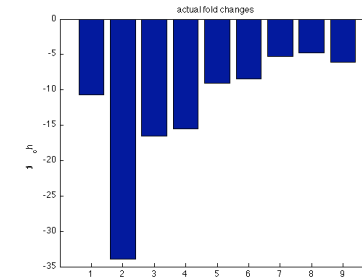
- Calculate average fold change for a group (e.g. HG vs. Normal)
- Select literature supported (143) and fold change based discriminators (i.e. with heatmap)

MicroRNA Signature Distinguishing HG/LG Bladder Cancer From Normal

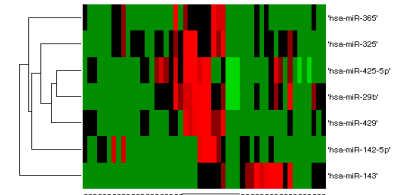
High Grade Bladder Cancer



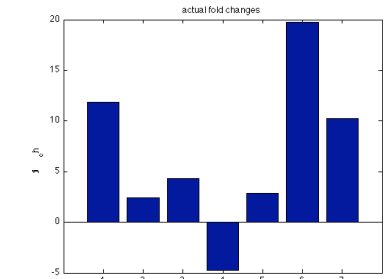
miR	fold change	P < 0.05
'hsa-miR-196b'	-10.7424526	
'hsa-miR-143'	-33.9199892	
'hsa-miR-23b'	-16.5776332	
'hsa-miR-30a-3p'	-15.5383422	
'hsa-miR-302a*'	-9.0335411	
'hsa-miR-329'	-8.46341989	
'hsa-miR-339'	-5.30631439	
'hsa-miR-373*'	-4.80447516	
'hsa-miR-496'	-6.05300353	



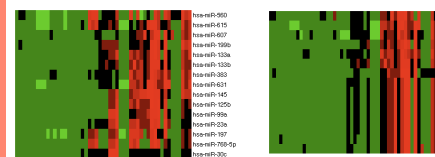
Low Grade Bladder Cancer



miR	fold	P < 0.05
'hsa-miR-142'	11.8381426	
'hsa-miR-29b'	2.44437238	
'hsa-miR-325'	4.32463512	
'hsa-miR-143'	-4.73075626	
'hsa-miR-425'	2.84153053	
'hsa-miR-365'	19.7338189	
'hsa-miR-429'	10.2700075	

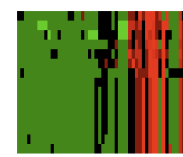


Bladder cancer signature



miR	Fold Change	P < 0.001
'hsa-miR-631'	-8.6832644	
'hsa-miR-145'	-46.5856007	
'hsa-miR-133a'	-19.0230341	
'hsa-miR-383'	-8.75437978	
'hsa-miR-23b'	-18.8986399	
'hsa-miR-125b'	-44.173594	
'hsa-miR-197'	-5.5230178	
'hsa-miR-123b'	-16.5249883	
'hsa-miR-99a'	-46.2086429	
'hsa-miR-30c'	-5.4995137	
'hsa-miR-607'	-10.3710262	
'hsa-miR-615'	-5.3064239	
'hsa-miR-768-5p'	-12.0061793	
'hsa-miR-569'	-4.38513412	
'hsa-miR-199b'	-8.9891183	

HG



miR	Fold Change	P < 0.001
'hsa-miR-631'	-4.0213824	
'hsa-miR-383'	-3.5647789	
'hsa-miR-607'	-6.68154368	
'hsa-miR-129'	-3.4240449	
'hsa-miR-190'	-5.3752055	
'hsa-miR-568'	-5.3752055	
'hsa-miR-568'	-5.3752055	
'hsa-miR-577'	-5.3752055	
'hsa-miR-630'	-5.3752055	
'hsa-miR-595'	-4.4018418	
'hsa-miR-362'	-6.4591282	
'hsa-miR-153'	-4.5855089	
'hsa-miR-133b'	-3.6478088	
'hsa-miR-214'	-6.16486656	

LG

Conclusion

- Top 15 differentially expressed microRNAs are down regulated in HG and LG bladder cancer.
- 9 MicroRNA signature set can distinguish HG bladder cancer from normal tissue, while 7 microRNAs can distinguish LG from normal
- Mir143 is common to both
- Further validation on more samples are needed.

References

Livak, K. J., & Schmittgen, T. D. (2001). Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the 2- $\Delta\Delta$ [Delta]Ct Method. *Methods*, 25(4), 402-408.

Schmittgen, T. D., & Livak, K. J. (2008). Analyzing real-time PCR data by the comparative Ct method. *Nat. Protocols*, 3(6), 1101-1108

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