

# BNORC Transgenic Core



## Mouse Genetic Engineering made “Easi”

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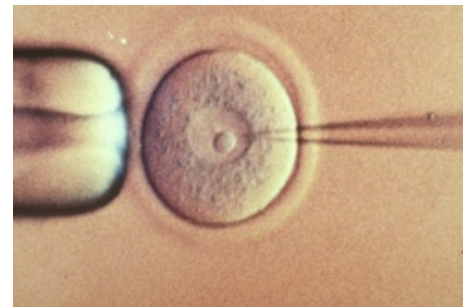
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# Two general types of mouse genetic engineering

## 1. Addition of exogenous DNA to the mouse genome – a.k.a. “Transgenics”.

Transgene

Promoter that Drives Expression

Protein Coding Sequence

**Trans-  
gene**

Ubiquitously active promoters  
Tissue-specific promoters:  
white fat, brown fat,  
muscle,  $\beta$ -cell, etc.

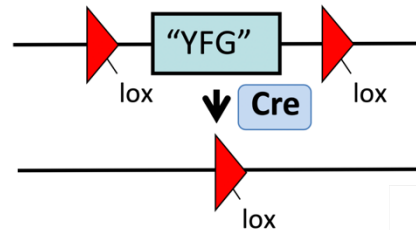
Anything  
Constitutively active,  
Dominant negative,  
etc., etc., etc.



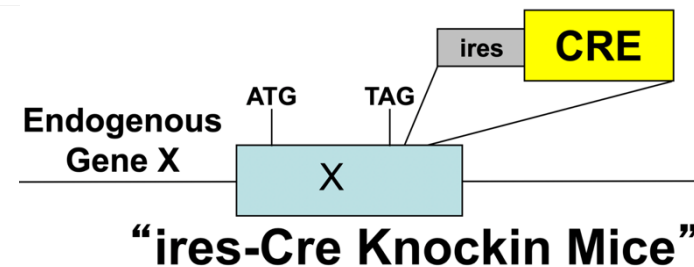
## 2. Targeted manipulation of the mouse genome – a.k.a. “Gene Targeting”.

- a) Endogenous gene is made nonfunctional – “global gene knockout”
- b) Endogenous gene is subtly altered – i.e. mimic a human point mutation, etc.

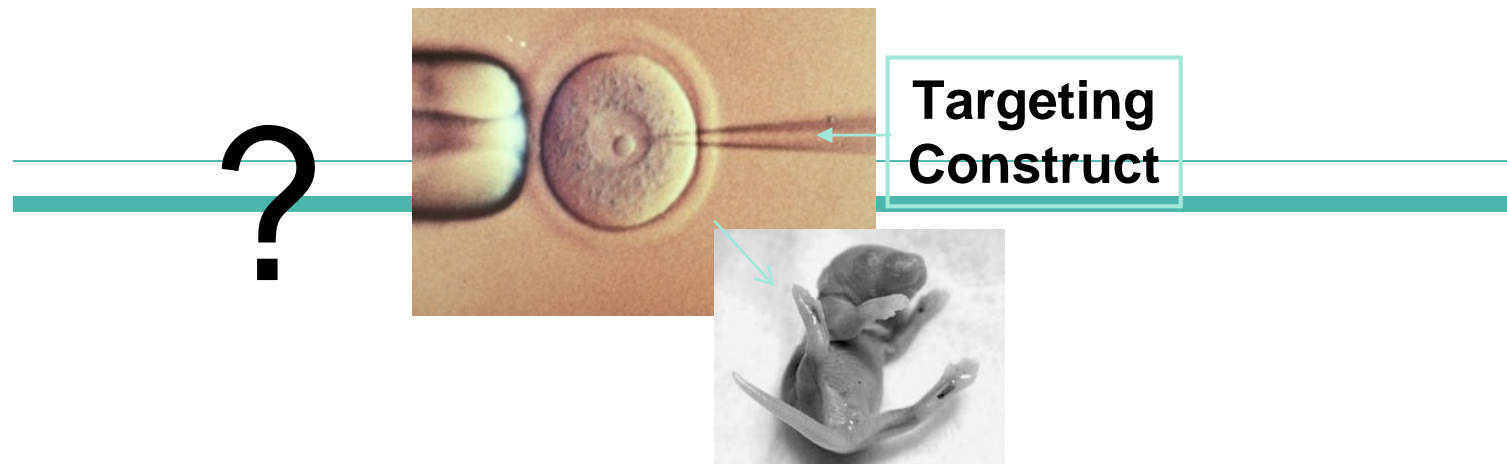
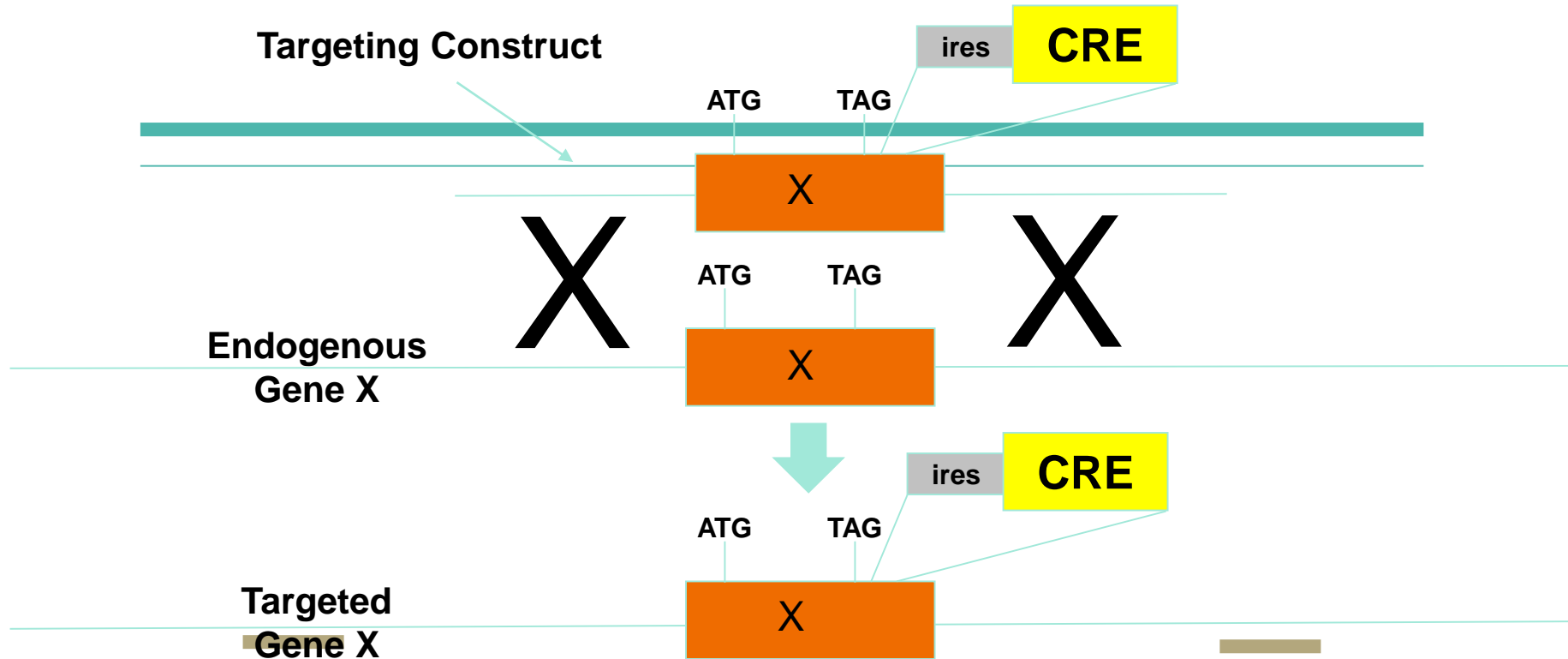
- c) Endogenous gene is “loxed” for doing tissue-specific knockouts

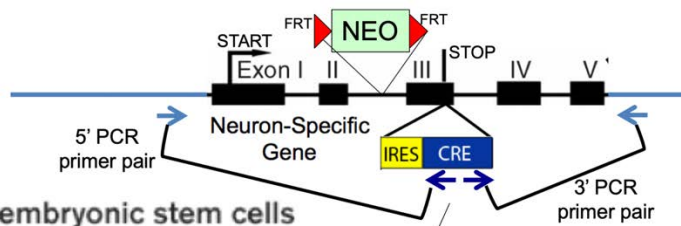


- d) Endogenous gene is modified to drive expression of a protein (anything): i.e. Cre Recombinase

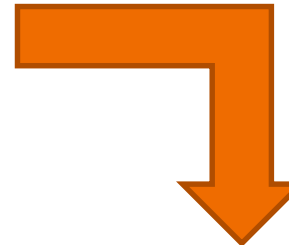
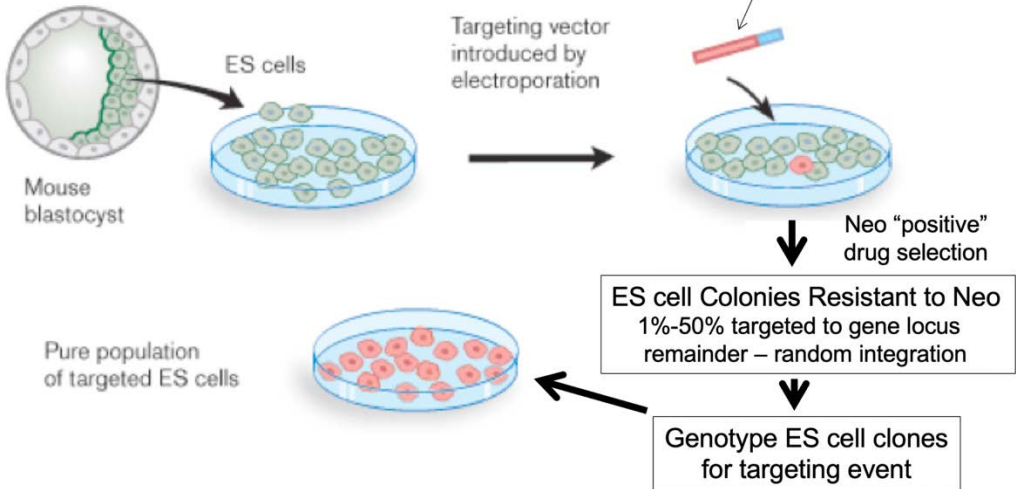


# Gene Targeting – key role of Homologous Recombination

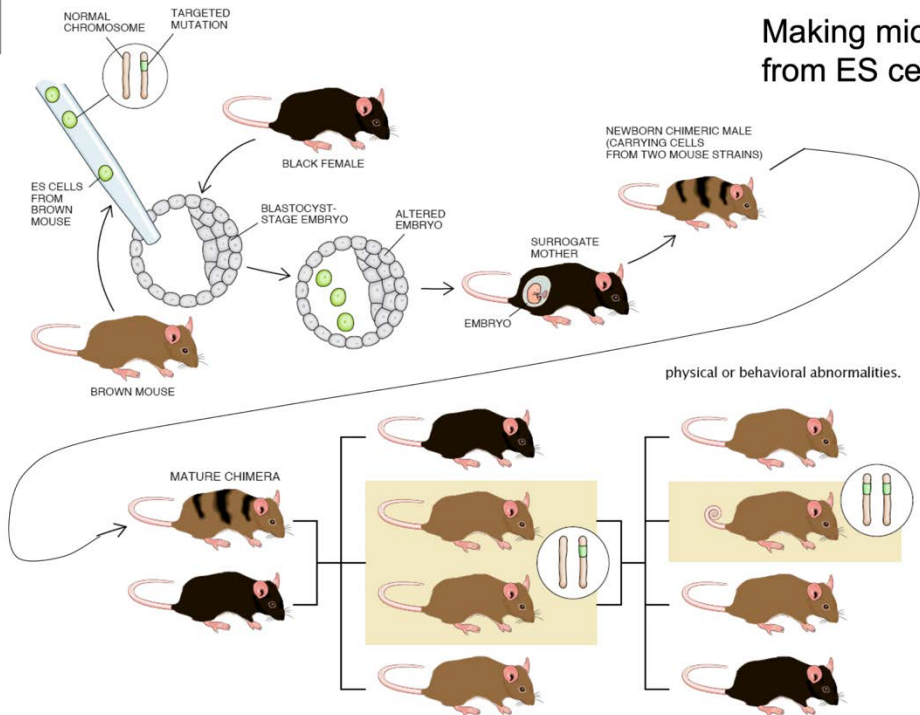




## A. Gene targeting of embryonic stem cells



## Making mice from ES cells



## The Nobel Prize in Physiology or Medicine 2007

"for their discoveries of principles for introducing specific gene modifications in mice by the use of embryonic stem cells"



Photo: U. Montan

Mario R. Capecchi



Photo: U. Montan

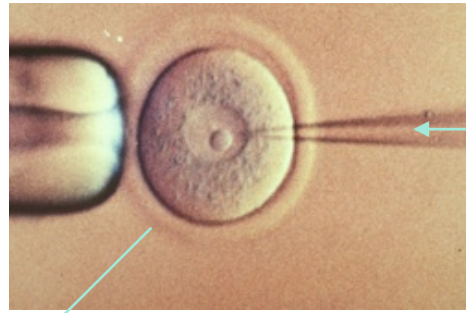
Sir Martin J. Evans



Photo: U. Montan

Oliver Smithies

# *Easi*-CRISPR: rapid, easy Gene Targeting



Targeting  
Construct  
and  
CRISPR  
"reagents"

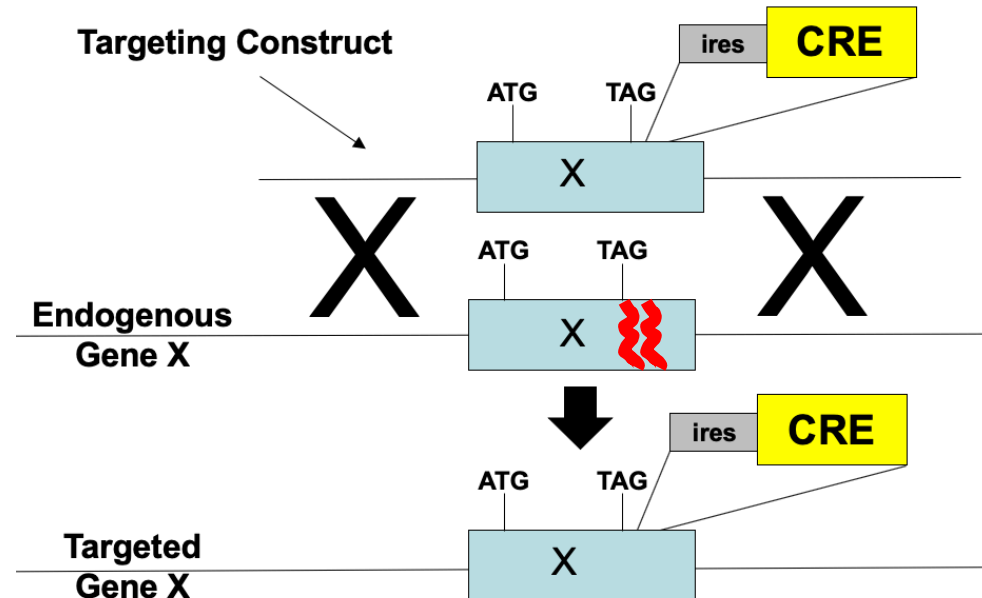


1. **CRISPR** is used to create **break** in DNA:

↑'s rate of homologous recombination (HR).

2. A **ssDNA** targeting construct is used:

- no random integration.
- ↑'s rate of HR.



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# Easy mouse gene targeting with Easi-CRISPR

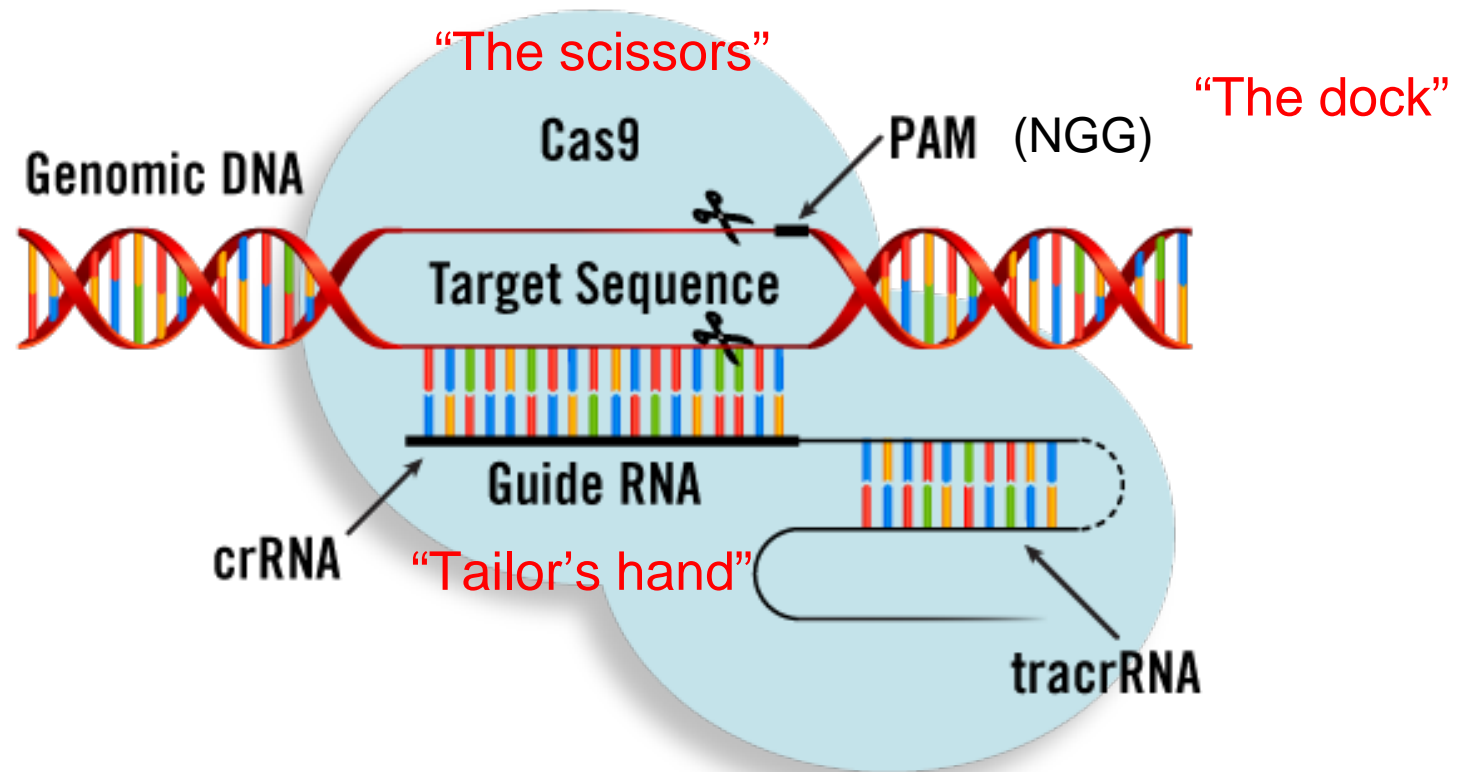
Chen Wu, PhD  
Co-director, BNORC Transgenic core  
Instructor in Medicine, BIDMC  
[cwu9@bidmc.harvard.edu](mailto:cwu9@bidmc.harvard.edu), 617-735-3259

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# An overview of CRISPR technology

# A brief overview of CRISPR technology

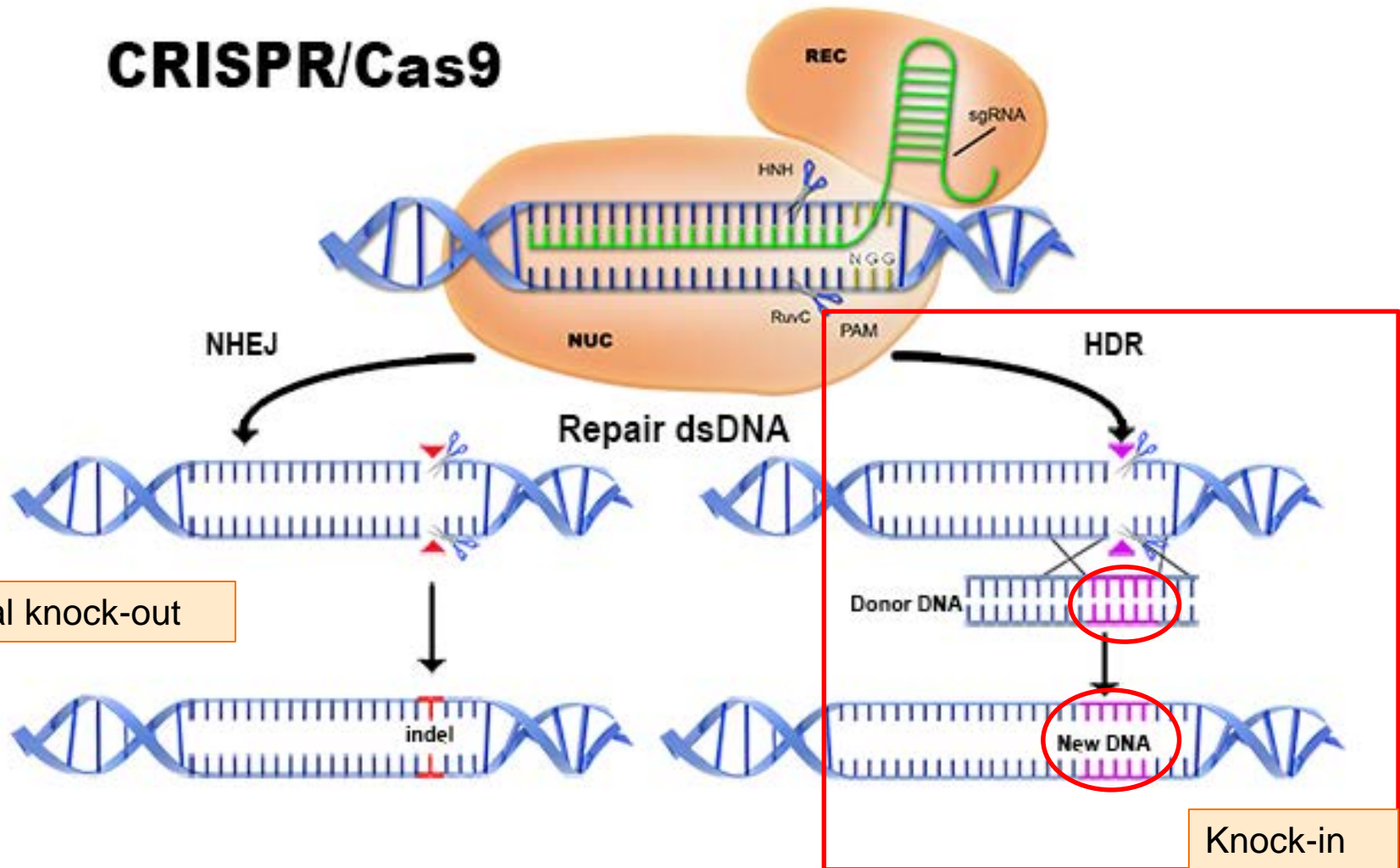
To create double-stranded DNA break





# Working with DSB

## CRISPR/Cas9

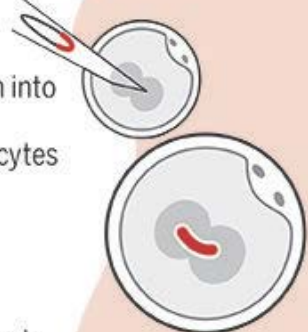


# CRISPR-Cas9

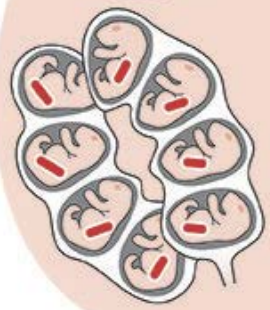
1 Order or design and produce guide RNAs



2 Microinjection into fertilized, one-celled oocytes



3 Transfer to pseudopregnant females



4 Birth of litter



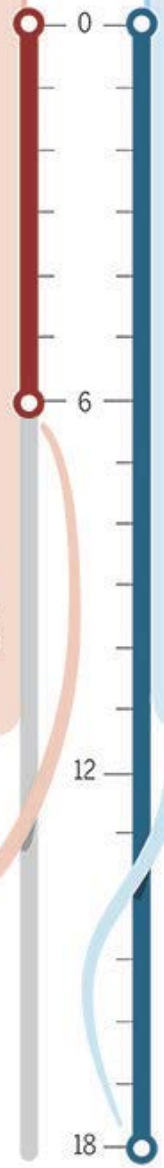
5 Skip chimeric stage

6 Germ line transmission occurs in all offspring

7 Expansion



Average time (months)

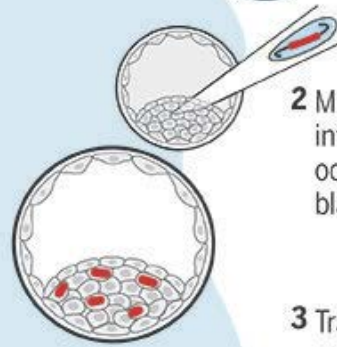


# Embryonic stem (ES) cell

1 Order or design and produce modified ES cells



2 Microinjection into fertilized oocytes in blastocyst stage



3 Transfer to pseudopregnant females



4 Birth of litter



5 Breed chimeric offspring with wild-type mice

6 Screen for germ line transmission

7 Expansion



CRISPR/CAS9 method greatly shortens the timeline

Easi-CRISPR

RESEARCH

Open Access



# *Easi*-CRISPR: a robust method for one-step generation of mice carrying conditional and insertion alleles using long ssDNA donors and CRISPR ribonucleoproteins

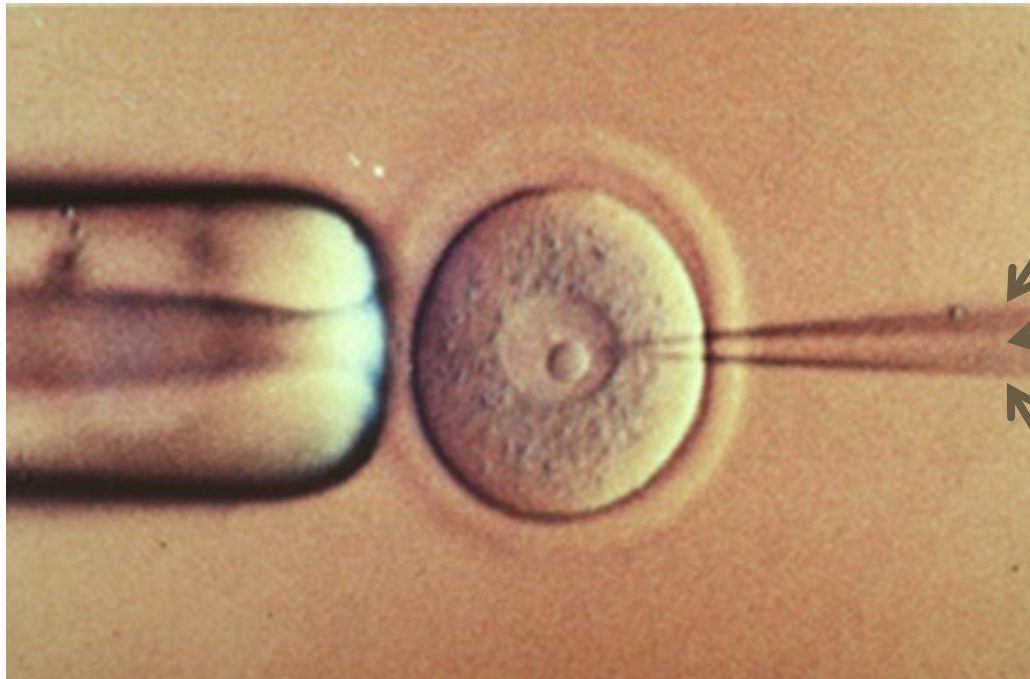
Rolen M. Quadros<sup>1†</sup>, Hiromi Miura<sup>2,3†</sup>, Donald W. Harms<sup>1</sup>, Hisako Akatsuka<sup>2,4</sup>, Takehito Sato<sup>4</sup>, Tomomi Aida<sup>5,6,7</sup>, Ronald Redder<sup>8</sup>, Guy P. Richardson<sup>9</sup>, Yutaka Inagaki<sup>3,10,11</sup>, Daisuke Sakai<sup>10,12</sup>, Shannon M. Buckley<sup>13,15</sup>, Parthasarathy Seshacharyulu<sup>14</sup>, Surinder K. Batra<sup>14,15</sup>, Mark A. Behlke<sup>16</sup>, Sarah A. Zeiner<sup>16</sup>, Ashley M. Jacobi<sup>16</sup>, Yayoi Izu<sup>17</sup>, Wallace B. Thoreson<sup>18</sup>, Lisa D. Urness<sup>19</sup>, Suzanne L. Mansour<sup>19\*</sup>, Masato Ohtsuka<sup>2,3,10\*</sup> and Channabasavaiah B. Gurumurthy<sup>1,20\*</sup>

# Comparison with conventional CRISPR

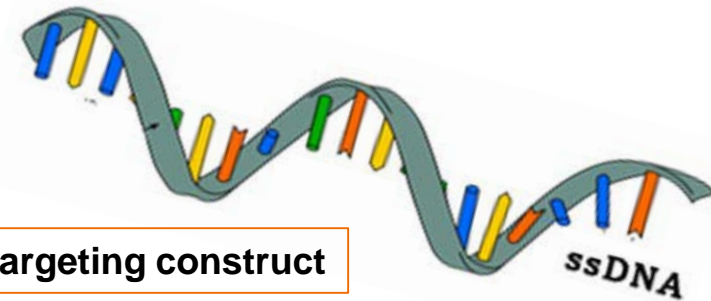
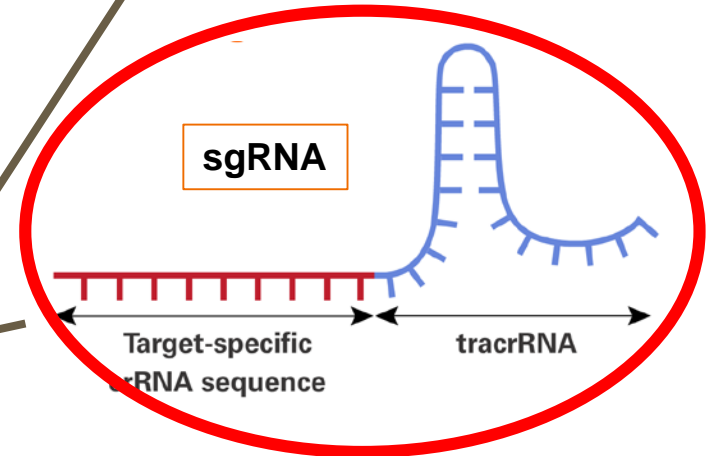
	Conventional	Easi-CRISPR
Targeting construct form	Plasmid (dsDNA)	ssDNA
Targeting construct availability	Cloning	Chemical synthesis
Homology arm length	> 2.5 kb	100 nt
Genotyping	Long range PCR / Southern blot (difficult)	Regular PCR (easy)
Success rate	< 10%	25% (and counting)



# Easi-CRISPR Reagents



CAS9 protein



Targeting construct

# sgRNA design: in silico

- Institutional sites: Broad, MIT, crispor.org
- Commercial sites: IDT, Synthego, etc.
- Cloud-based bioinformatics sites: Benchling.com

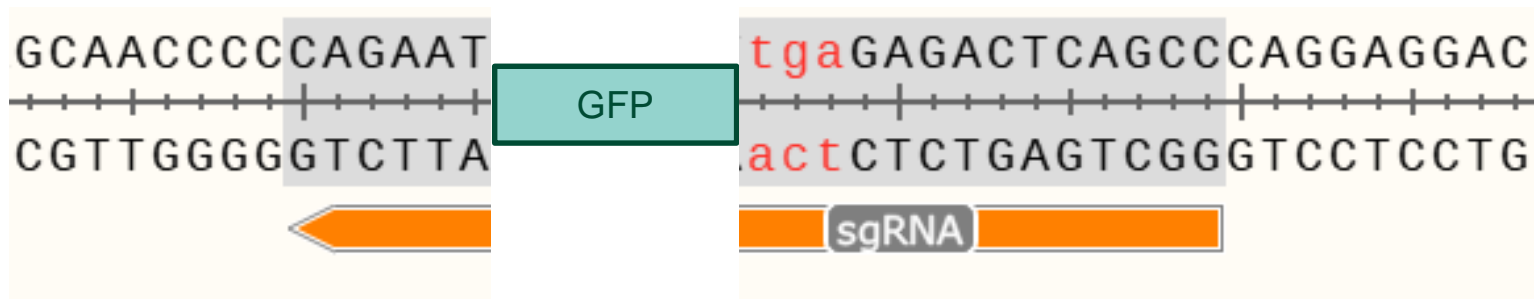
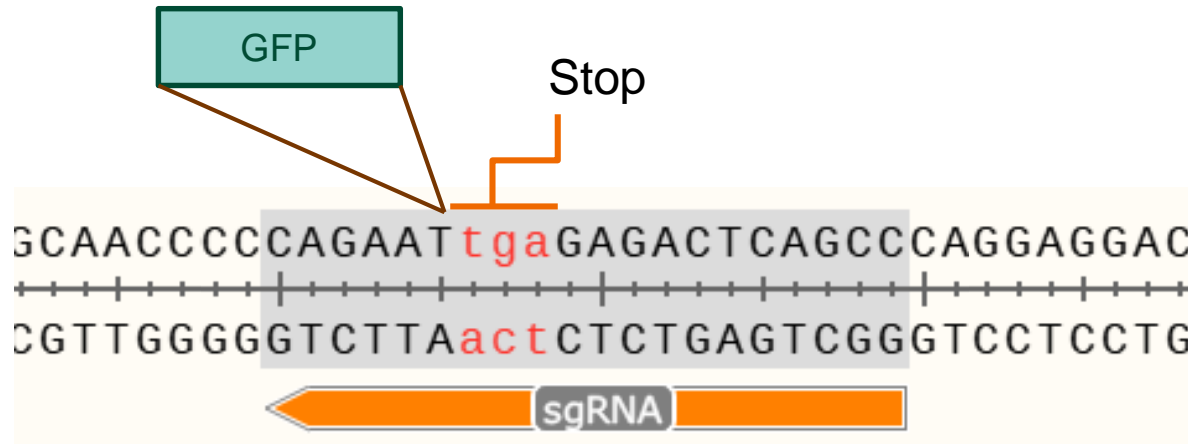
Online predicting algorithms are based on analysis of 3000+ sgRNAs (Doench et al. 2014 and 2016)

# Criteria of a good sgRNA

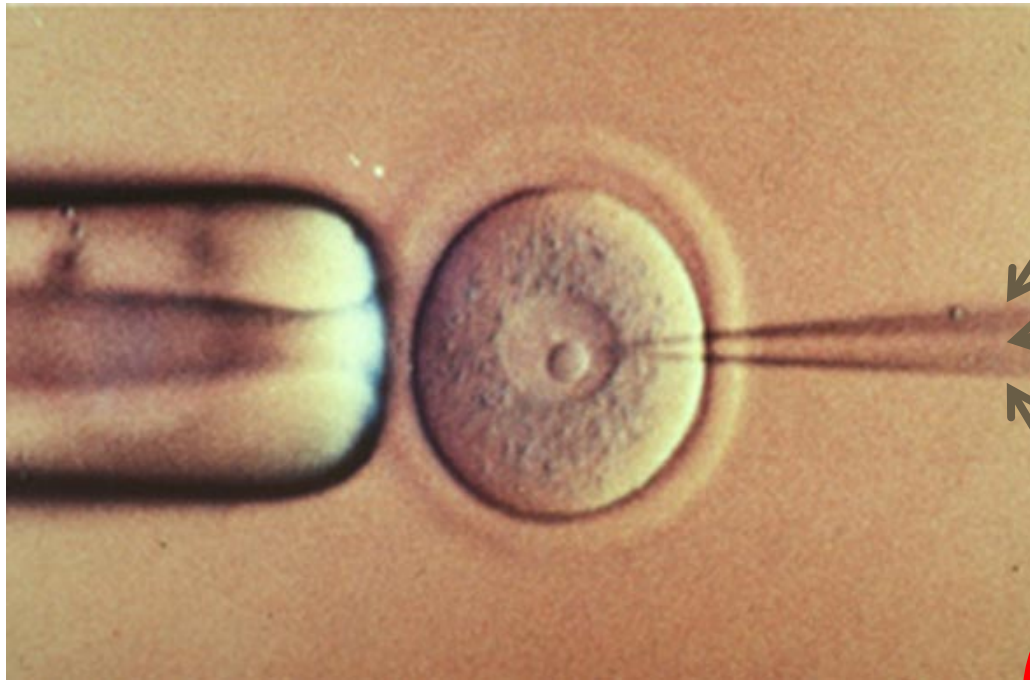
- Efficacy or on-target score – cuts where it should
- Specificity or off-target score – does not cut where it shouldn't
- Close to the insertion site



# The ideal sgRNA site: contains the insertion site

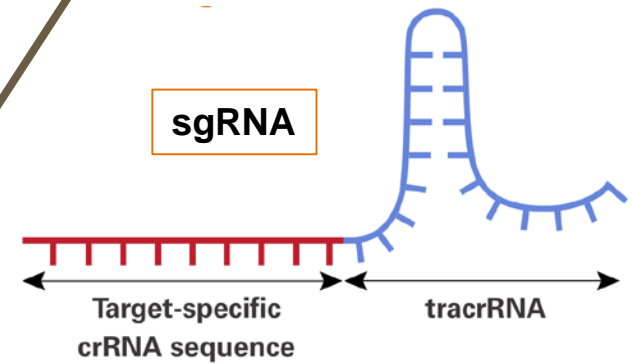


# Easi-CRISPR Reagents



CAS9 protein

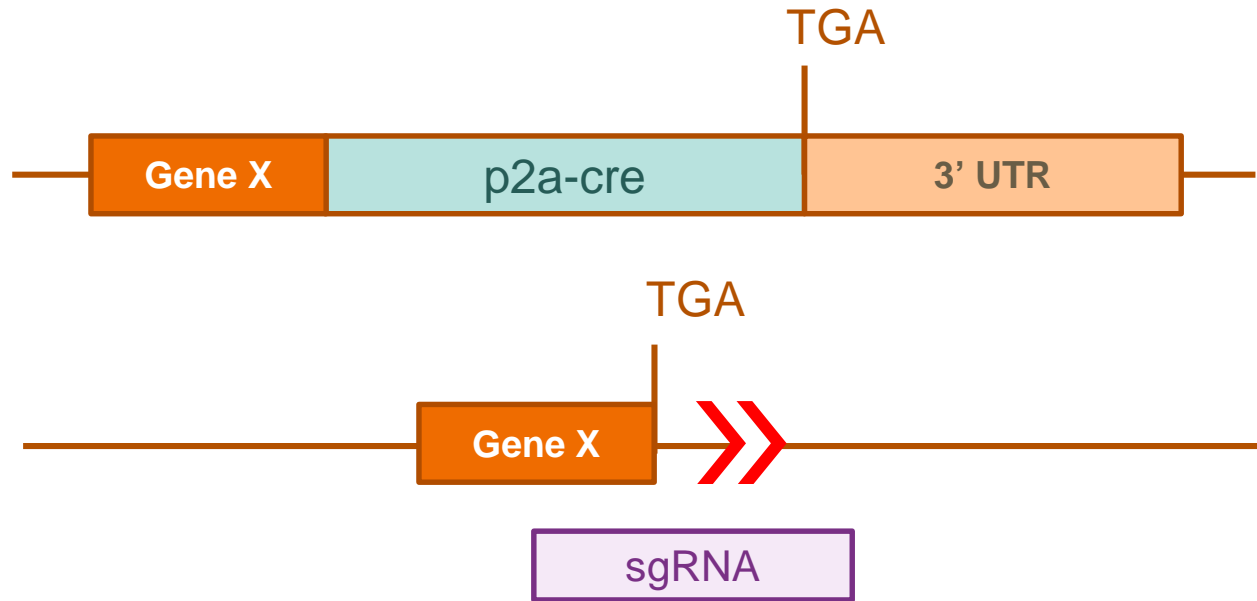
sgRNA



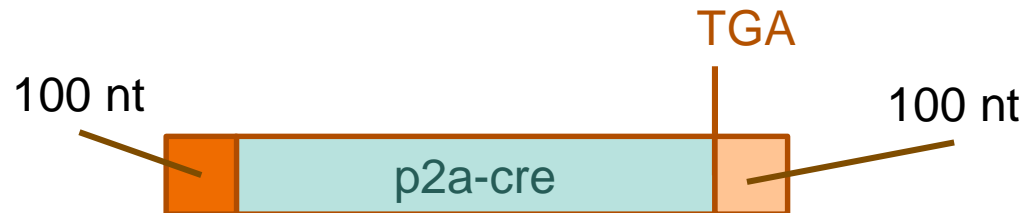
Targeting construct



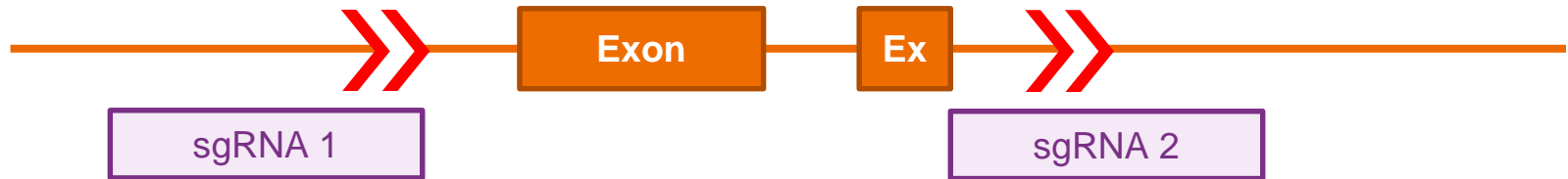
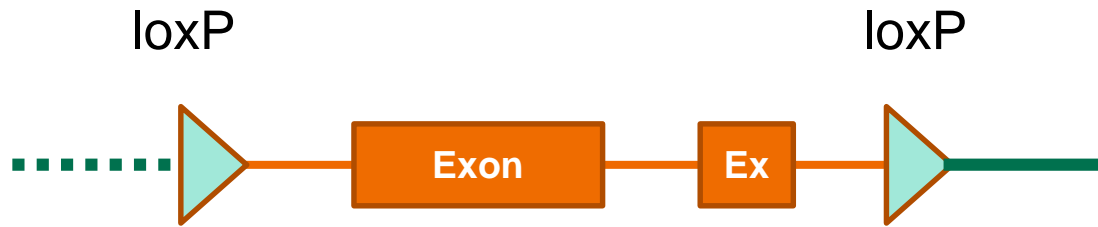
# Project design (I) – reporter or driver



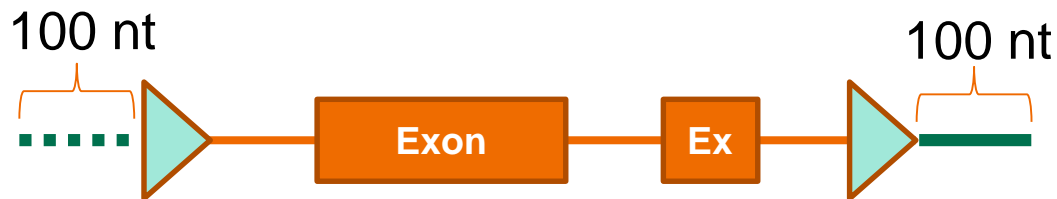
Targeting Construct:



# Project design (II) – floxed exon(s)



Targeting construct:



# Easi-CRISPR limitation: template availability

1. size: 2kb (IDT), 5kb (Genewiz)
2. Sequence composition: GC content, repetitive sequence, etc.
3. Cost: 90 cents/nt
4. Time: ~ 1 month

# Easi-CRISPR Workflow

1. Contact us
  2. Design sgRNA and targeting construct
  3. Order the reagents
  4. Send the reagents to us
- 4 weeks later...
5. Pick up the tail snips and genotype
  6. Breed the founder(s)

# Project summary

# Summary

- Turn-around time: 3 months or sooner (from reagents to founder)
- Average success rate (Easi-CRISPR): 25% (71% the highest)
- Number of successful projects : 50+ within 2.5 years
- Including 4 co-injections: two targeting constructs with one sgRNA
- **We provide free consultation on strategy and design**



Questions and comments?

Thank you!!



Joel Lawitts, PhD  
Director, Transgenic core